

FORM PTO-1390 (Modified)
(REV 11-2000)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

209956US0XPCT

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

09/857179

INTERNATIONAL APPLICATION NO.
PCT/JP00/07216INTERNATIONAL FILING DATE
18 OCTOBER 2000PRIORITY DATE CLAIMED
19 OCTOBER 1999

TITLE OF INVENTION

ANTIURACIL MONOCLONAL ANTIBODY

APPLICANT(S) FOR DO/EO/US

Gen-ichiro SOMA

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below.
4. ☐ The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
 - a. ☐ is attached hereto (required only if not communicated by the International Bureau).
 - b. ☒ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
 - a. ☒ is attached hereto.
 - b. ☐ has been previously submitted under 35 U.S.C. 154(d)(4).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
 - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
10. ☐ An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).
11. ☐ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☒ A copy of the International Search Report (PCT/ISA/210).

Items 13 to 20 below concern document(s) or information included:

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☐ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☐ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
20. ☐ A second copy of the published international application under 35 U.S.C. 154(d)(4).
21. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
22. ☐ Certificate of Mailing by Express Mail
23. ☒ Other items or information:

Request for Consideration of Documents in International Search Report
 Notice of Priority / PCT/IB/304 / PCT/IB/308
 Drawings (2 sheets)

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.53)

097857179

INTERNATIONAL APPLICATION NO.

PCT/JP00/07216

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209956US0XPCT

24. The following fees are submitted:

BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :

- ☐ Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1000.00
- ☒ International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$860.00
- ☐ International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$710.00
- ☐ International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$690.00
- ☐ International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00

ENTER APPROPRIATE BASIC FEE AMOUNT =**\$860.00**

Surcharge of **\$130.00** for furnishing the oath or declaration later than ☐ 20 ☐ 30 months from the earliest claimed priority date (37 CFR 1.492 (e)).

\$0.00

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE
Total claims	12 - 20 =	0	x \$18.00
Independent claims	1 - 3 =	0	x \$80.00
Multiple Dependent Claims (check if applicable).			<input checked="" type="checkbox"/>

\$270.00**TOTAL OF ABOVE CALCULATIONS =****\$1,130.00**

- ☐ Applicant claims small entity status. (See 37 CFR 1.27). The fees indicated above are reduced by 1/2.

\$0.00**SUBTOTAL =****\$1,130.00**

Processing fee of **\$130.00** for furnishing the English translation later than ☐ 20 ☐ 30 months from the earliest claimed priority date (37 CFR 1.492 (f)).

\$0.00**TOTAL NATIONAL FEE =****\$1,130.00**

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable).

\$0.00**TOTAL FEES ENCLOSED =****\$1,130.00**Amount to be:
refunded

\$

charged

\$

- a. ☒ A check in the amount of **\$1,130.00** to cover the above fees is enclosed.
- b. ☐ Please charge my Deposit Account No. _____ in the amount of _____ to cover the above fees. A duplicate copy of this sheet is enclosed.
- c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. **15-0030**. A duplicate copy of this sheet is enclosed.
- d. ☐ Fees are to be charged to a credit card. **WARNING:** Information on this form may become public. **Credit card information should not be included on this form.** Provide credit card information and authorization on PTO-2038.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

**22850**

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REGISTRATION NUMBER

6-19-01

DATE

2/PRTS

09/857179
JC18 Rec'd PCT/PTO 1 9 JUN 2001

Description

Antiuracil Monoclonal Antibody

Technical Field

The present invention relates to a monoclonal antibody useful for screening DPD-deficient cancer patients for whom administration of a pyrimidine fluoride-based antitumor agent is contraindicated; a hybridoma producing the monoclonal antibody; a method of immunochemically assaying uracil or thymine by use of the monoclonal antibody; and a diagnostic agent containing the monoclonal antibody for diagnosing DPD deficiency.

Background Art

As antitumor agents, pyrimidine fluoride-based compounds such as fluorouracil are currently used. However, among cancer patients, some (approximately 3% of Caucasian breast cancer patients) are inherently deficient in dihydropyrimidine dehydrogenase (hereinafter abbreviated as DPD), which is an enzyme involved in the metabolism of the compound in its decomposition pathway. It has been reported that, when a pyrimidine fluoride-based antitumor agent is administered to such a DPD-deficient patient, the compound is not metabolized and remains in the body, leading to side effects or possibly even to death (*Biochim. Biophys. Acta* 633, 400-409 (1980)).

In an organism, uracil and thymine are generally metabolized into dihydrouracil and dihydrothymine, respectively, through the action of DPD. However, DPD-deficient patients are known to fail to metabolize uracil and thymine and to excrete large amounts of unchanged uracil and thymine into the blood or urine, particularly uracil (*Adv. Exp. Med. Biol.*, 253A, 111-118 (1989)). Accordingly, if uracil or thymine in blood or urine is assayed prior to administration of a pyrimidine fluoride-bases antitumor agent to cancer patients, DPD-deficient patients can be screened in advance. As a result, use of the antitumor agent can be discontinued or the dose of the agent can be reduced, to thereby avoid serious side effects.

Conventionally, methods for assaying uracil have been known, including a high-performance liquid chromatographic method (*Journal of Chromatography B*, 672 (1995), 233-239) and an immunoassay method making use of a monoclonal antibody for pseudouridine (Japanese Patent Publication (*kokoku*) 4-21479). However, the former method is disadvantageous in that the preparation of samples is cumbersome, imposes a heavy workload, and requires a significant level of skill. In addition, a long period of time is generally required for assaying a number of samples, and the high cost of measurement and other apparatuses make the method disadvantageous. In the latter method, the employed monoclonal antibody—an antibody used for diagnosing a progressive cancer—exhibits a reactivity with uracil of

30-40% and also a reactivity with pseudouridine as high as 95-99%. Thus, use of such a monoclonal antibody that has cross-reactivity with uracil and pseudouridine cannot distinguish DPD-deficient patients from non-deficient patients.

Under such circumstances, the present inventors previously found a monoclonal antibody which reacts strongly with uracil without cross-reacting with dihydrouracil—a metabolite of uracil in the organism—and with pseudouridine employed as a tumor marker (International Patent Publication WO 99/20748).

However, the above monoclonal antibody also reacts strongly with N-carbamyl- β -alanine, which is a metabolite of dihydrouracil and is contained in large amounts in normal urine, and has insufficient reactivity with uracil. Thus, use of the monoclonal antibody as a diagnostic agent for DPD deficiency is problematic.

Thus, an object of the present invention is to provide a monoclonal antibody having a higher specificity with uracil and thymine. Another object is to provide a hybridoma producing the monoclonal antibody. Still another object is to provide an immunochemical assay method which can correctly determine uracil and thymine contained in urine. Yet another object is to provide a diagnostic agent containing the monoclonal antibody for diagnosing DPD deficiency.

Disclosure of the Invention

In view of the foregoing, the present inventors have carried out extensive studies on the relevant immunogen and immunization methods, and have found that a monoclonal antibody produced from a hybridoma which is formed by administering 5-bromo-1-carboxymethyluracil to an animal strongly reacts specifically with both uracil and thymine but exhibits no or low reactivity with pseudouridine, dihydrouracil, dihydrothymine, and N-carbamyl- β -alanine, and is therefore very useful for diagnosing DPD deficiency. The present invention has been accomplished on the basis of this finding.

Accordingly, the present invention provides a monoclonal antibody, characterized by reacting strongly with uracil and thymine but scarcely with N-carbamyl- β -alanine.

The present invention also provides a hybridoma producing the monoclonal antibody.

The present invention also provides a method for immunochemically assaying uracil and/or thymine by use of the monoclonal antibody.

The present invention also provides a diagnostic agent for diagnosing DPD deficiency, which agent contains the monoclonal antibody.

The present invention also provides a method for diagnosing DPD deficiency, characterized by assaying uracil and thymine in a specimen by use of the diagnostic agent.

The present invention also provides use of the monoclonal antibody for producing a diagnostic agent for

diagnosing DPD deficiency.

The monoclonal antibody of the present invention reacts selectively with uracil and thymine at high sensitivity, to thereby selectively assay, rapidly and simply, uracil and thymine contained in a sample. In addition, the monoclonal antibody, exhibiting no or low reactivity with pseudouridine, dihydrouracil, and N-carbamyl- β -alanine, is useful for diagnosing DPD deficiency by use of a human urine sample. Particularly, the monoclonal antibody is considerably useful for screening DPD-deficient cancer patients for whom administration of a pyrimidine fluoride-based antitumor agent is contraindicated.

Brief Description of the Drawings

Fig. 1 is a graph showing reactivity with uracil for the monoclonal antibody of the present invention (CMU-1) and SU-1 studied through indirect competitive inhibition ELISA (Example 8).

Fig. 2 is a graph showing the relationship between the percent absorption of the color-developed monoclonal antibody of the present invention and the concentrations of uracil, thymine, and pseudouridine.

Best Modes for Carrying Out the Invention

The monoclonal antibody of the present invention reacts strongly with uracil and thymine but scarcely with N-carbamyl- β -alanine. The expression "scarcely with" refers to

a state that the monoclonal antibody exhibits no dose-dependent response in connection with N-carbamyl- β -alanine.

Preferably, the monoclonal antibody of the present invention reacts strongly with uracil and thymine but scarcely with N-carbamyl- β -alanine, and exhibits no or low reactivity with pseudouridine, dihydrouracil, and dihydrothymine. The expression "no or low reactivity" refers to a state that no dose-dependent response is observed in connection with a specific compound concentration or to a state that a dose-dependent response is observed in connection with a specific compound concentration but affinity to the compound is low at a specific absorbance.

More specifically, the monoclonal antibody preferably exhibits a selectivity in cross reaction with N-carbamyl- β -alanine of 10% or less, when the selectivity in cross reaction with uracil or thymine is 90% or more. More preferably, when the selectivity in cross reaction with uracil or thymine is 90% or more, the monoclonal antibody exhibits a selectivity in cross reaction with N-carbamyl- β -alanine of 10% or less; a selectivity in cross reaction with pseudouridine of 33% or less; a selectivity in cross reaction with dihydrouracil of 8% or less; and a selectivity in cross reaction with dihydrothymine of 23% or less.

The globulin type of the monoclonal antibody of the present invention is not particularly limited, and any monoclonal antibody is acceptable so long as the antibody reacts strongly with uracil and/or thymine but scarcely with

N-carbamyl- β -alanine. Specifically, any of IgG, IgM, IgA, IgE, and IgD is acceptable, with IgG and IgM being preferred.

No particular limitation is imposed on the cell strain which produces the monoclonal antibody of the present invention, so long as the produced antibody is endowed with the aforementioned characteristics. Examples of preferred cell strains include a hybridoma which is obtained through cell fusion of an antibody-producing cell with a myeloma cell strain.

As an antibody-producing cell for obtaining the monoclonal antibody of the present invention, there can be used a spleen cell, a lymphonodus cell, or a B-lymphocyte of an animal which is immunized *in vivo* with 5-halogeno-1-carboxymethyluracil serving as an immunogen. Examples of a halogen atom occupying the 5-position of 5-halogeno-1-carboxymethyluracil include fluorine, chlorine, bromine, and iodine, with a bromine atom being preferred.

The 5-halogeno-1-carboxymethyluracil *per se* has considerably weak immunogenicity. Thus, during immunization, the compound is preferably linked to an appropriate Schlepper, to thereby form an immunogen-Schlepper complex serving as an immunogen.

The term "Schlepper" refers to a carrier which links to a substance of very weak immunogenicity or to a substance, such as hapten, which *per se* has no antibody-productivity, to thereby potentiate or express immunogenicity. Generally, a protein of relatively high molecular weight is employed as a

Schlepper. In addition to such a protein, cells such as erythrocyte and polysaccharide may also be used. Examples of the Schlepper include hemocyanine from SUKASHI-GAI (KLH), ovalbumin (OVA), bovine serum albumin (BSA), and rabbit serum albumin, with KLH and BSA being particularly preferred.

The method of linking 5-bromo-1-carboxymethyluracil and Schlepper is not particularly limited, and any known method can be employed. Examples include an acid anhydride mixture method (B. F. Erlanger *et al.*, *J. Biol. Chem.* 234 1090-1094 (1954)) and an activation ester method (A. B. KARU *et al.*, *J. Agric. Food. Chem.* 423-309 (1994)). There may also be employed, as a simple technique, a method employing an Immject Immunogen EDC Conjugation kit (product of Pierce) in accordance with the manual attached thereto.

Animals to be immunized include mice, rats, horses, goats, and rabbits, and an antigen is administered to these animals through a routine method. Specifically, an adjuvant such as complete Freund's adjuvant or incomplete Freund's adjuvant and the aforementioned 5-halogeno-1-carboxymethyluracil-KLH or 5-halogeno-1-carboxymethyluracil-BSA are mixed to prepare a suspension or emulsion thereof, and the suspension or emulsion is administered intravenously, subcutaneously, intradermally, intraperitoneally, or in a similar manner, into an animal, such as a mouse, several times so as to immunize the animal.

An antibody-producing cell such as a spleen cell is obtained from the thus-immunized animal and fused with a

myeloma cell, to thereby produce a hybridoma according to the present invention.

Although the myeloma cell strains to be fused can be derived from mice, rats, horses, goats, rabbits or humans, preferably the cell strains are derived from the same animal from which an antibody-producing cell is obtained. For example, when an antibody-producing cell is derived from a mouse spleen cell, a counter myeloma cell strain obtained from a mouse is preferably used as the cell to be fused. Specifically, the myeloma cell strain is an 8-azaguanine-resistant mouse (BALB/c-derived) myeloma cell strain. Examples of specific cells include P3/X63-Ag8 (X63) [*Nature*, 256, 495-497 (1975)], P3/X63-Ag8.U1 (P3U1) [*Current Topics in Microbiology and Immunology*, 81, 1-7 (1987)], P3/NSI-1-Ag4-1 (NS-1) [*Eur. J. Immuno Meth.*, 35, 1-21 (1976)], Sp2/O-Ag14 (Sp2/O) [*Nature* 276, 269-270 (1978)], FO [*J. Immuno Meth.*, 35, 1-21 (1980)], MPC-11, X63.653, and S194, with P3U1 being preferred.

Cell fusion of an antibody-producing cell with a myeloma cell can be performed through any known method (e.g., see *Nature*, 256, 495-497 (1975), *Proc. Natl. Acad. Sci. USA* 78, 5122-5126 (1981)) or a method similar thereto. Specifically, the aforementioned antibody-producing cell and myeloma cell are fused in a conventionally employed nutrient medium in the presence of a fusion accelerator. The fusion accelerator is not particularly limited, and any of the typical accelerators, polyethylene glycol, Sendai virus, etc.

can be used (e.g., Shuji YAMASHITA et al., Cell Tissue Chemistry, edited by Nihon Soshiki Saibo Kagaku-Kai; Gakusai Kikaku, 1986). Of these, polyethylene glycol is preferred in view of comparatively low cytotoxicity and ease of the fusion operation. In addition, an optional adjuvant such as dimethyl sulfoxide may be used in combination so as to enhance the fusion efficiency. Instead of the fusion method employing the aforementioned fusion accelerator, a fusion method through electric treatment (electric fusion) may appropriately be employed.

The number ratio of antibody-producing cells to myeloma cells may fall within the range employed in routine methods. Specifically, the antibody-producing cells may be used in an amount approximately 2-10 times, preferably 4-7 times, that of the myeloma cells.

A hybridoma group which has been endowed with the ability of antibody-production and proliferation, through cell fusion, of the antibody-producing cell and the myeloma cell can be selected through culturing by use of a customary medium for selection. Examples of the medium for selection include HAT medium when an 8-azaguanine-resistant strain is employed as the myeloma cell strain. The culturing in the HAT medium may be performed for a sufficient period of time such that unfused cells and the like, other than the target hybridoma, are extirpated, typically over 3-10 days.

Subsequently, the thus-obtained hybridoma is subjected to screening for strains producing a target anti-uracil

monoclonal antibody and subsequent cloning, wherein the screening and cloning are carried out in accordance with a conventional method.

Screening strains which produce the anti-uracil monoclonal antibody of the present invention is performed by collecting a part of the culture supernatant containing the thus-obtained hybridoma group, to thereby prepare a sample, and detecting (confirming) uracil contained in the thus-collected sample through any of a variety of routine antibody-detection methods ("Hybridoma Method and Monoclonal Antibody," published by R&D Planning Co., Ltd., p. 30-53, March 5, 1982); e.g., radioimmunoassay. Examples of the antibody-detection method include ELISA, a fluorescent antibody technique, a plaque method, a spot method, hemagglutination, and the Ouchterlony method. Of these, ELISA, particularly indirect competitive inhibition ELISA, is preferably employed, in view of sensitivity, speedy operation, accuracy, safety, and automation.

The hybridoma which has been proven, by screening, to produce a uracil-specific monoclonal antibody is cloned through a method in which a cell liquid is diluted through limiting dilution such that one hybridoma is included in one well (limiting dilution); a method in which the cell liquid is spread onto a soft agar medium and the formed colony is collected; a method in which one cell is taken by means of a micromanipulator; or a method in which one cell is separated by means of a cell-sorter (sorter cloning).

As the cloning method, limiting dilution is preferably employed, in view of ease of operation. Specifically, wells containing a hybridoma for which antibody titer has been confirmed are subjected to limiting dilution 1-4 times, and the hybridoma for which antibody titer has been constantly confirmed is selected as a cell strain producing an anti-uracil monoclonal antibody.

The hybridoma of the present invention obtained through the aforementioned method may be stored by freezing at -80°C or lower (e.g., -195°C in liquid nitrogen) in a culture medium; e.g., RPMI 1640 medium containing 10% fetal bovine serum and including 5×10^6 cells/mL or more, with optional 10% dimethyl sulfoxide serving as a freeze stabilizing agent.

As further mentioned, the hybridoma of the present invention is stable during culture in a basal medium such as RPMI 1640, DMEM, or IMEM and produces and secretes a monoclonal antibody which reacts specifically with both uracil and thymine but exhibits low or scarce reactivity with pseudouridine, dihydrouracil, dihydrothymine, and N-carbamyl- β -alanine. In addition, the hybridoma, stabilized in nitrogen liquid, can be stored therein and can be readily reclaimed therefrom. The hybridoma of the present invention (mouse hybridoma CMU-1) is useful as a readily available source for producing a genuine monoclonal antibody reacting with a uracil antigen, and is deposited as "FERM BP-6870" (date of deposition: 1999/9/7) with the National Institute of Bioscience and Human-Technology, Agency of Industrial Science

and Technology, Ministry of International Trade and Industry
(address: 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken 305-
8566 JAPAN) under the Budapest Treaty.

The target monoclonal antibody is produced from the
thus-obtained hybridoma by culturing the hybridoma in a
routine manner such as cell culture or formation from
ascites; separating the monoclonal antibody from the culture
supernatant or the ascites; and purifying. The monoclonal
antibody obtained from the culture supernatant or ascites can
be purified through a routine method. Specifically, methods
such as fractionating by use of ammonium sulfate, gel
filtration, ion-exchange chromatography, and affinity
chromatography may appropriately be applied in combination.

The thus-obtained monoclonal antibody of the present
invention can be evaluated in terms of selectivity in cross-
reaction, through a method such as indirect competitive
inhibition ELISA as described in the Examples shown below.
In the Examples, the monoclonal antibody has been proven to
react specifically with both uracil and thymine and to
exhibit low or scarce reactivity with pseudouridine,
dihydrouracil, dihydrothymine, and N-carbamyl- β -alanine; i.e.,
has been proven to have high reactivity with uracil and
thymine and high reaction specificity thereto.

Thus, the monoclonal antibody of the present invention
has high reactivity with both uracil and thymine but exhibits
low or scarce reactivity with pseudouridine, dihydrouracil,
dihydrothymine, and N-carbamyl- β -alanine. In addition, the

antibody can be supplied constantly and in a large amount by culturing the hybridoma of the present invention. Therefore, the antibody is useful as a reagent for specifically assaying uracil and thymine immunochemically and can be used for purifying uracil and thymine.

As described above, the monoclonal antibody of the present invention has reaction specificity; i.e., low or scarce reactivity with pseudouridine, dihydrouracil, dihydrothymine, and N-carbamyl- β -alanine, which are substances that possibly migrate into the urine of cancer patients. Thus, the monoclonal antibody of the present invention is useful for screening DPD-deficient patients excreting uracil and thymine into the urine as a result of DPD deficiency and DPD-non-deficient patients, particularly for cancer patients.

No particular limitation is imposed on the method for immunochemically assaying uracil and thymine by use of the monoclonal antibody of the present invention, so long as the method can specifically detect or determine uracil and thymine in a sample possibly containing uracil and thymine. However, a preferred assay method includes a step of incubating a biological sample of a patient and the monoclonal antibody of the present invention under the conditions for forming an antigen-antibody complex, and a step of detecting the formed antigen-antibody complex. The above assay method may be varied in accordance with techniques that are typically employed in the art relating to

the present invention, and examples of the variations include direct, indirect, competitive, and sandwich methods. Specific examples include ELISA, radioimmunoassay, a fluorescent antibody technique, and an emission antibody technique, with ELISA being preferred.

In one example, immobilized ELISA of a human urine sample can be performed in the following manner:

(a) immobilizing the monoclonal antibody of the present invention onto an arbitrary carrier, and covering, with a protein irrelevant to the antigen, a part of the solid surface that is not covered with the antibody;

(b) washing the surface and adding the urine sample to be assayed and an enzyme-labeled antigen; e.g., enzyme-labeled (5-halogeno-1-carboxymethyluracil)-Schlepper complex so as to cause a competitive reaction; and (c) adding an enzyme substrate thereto and measuring the decrease in absorbance attributable to addition of the sample to be assayed. Briefly, detection of the decrease in absorbance attributable to addition of the sample to be tested can determine the presence of uracil in the urine sample, and measuring the amount of the decrease in absorption attributable to addition of the sample to be tested can quantitatively determine uracil and thymine in the urine sample, by use of a calibration curve established in advance from data on predetermined amounts of uracil and thymine.

Indirect competitive inhibition ELISA by use of the monoclonal antibody of the present invention can assay uracil

or thymine in a sample within a range of 0.001-2 mg/mL, preferably 0.005-0.5 mg/mL. Assaying uracil or thymine in a urine sample through indirect competitive inhibition ELISA can be performed in the following manner.

(a) causing a (5-halogeno-1-carboxymethyluracil)-Schlepper complex serving as an antigen to adsorb onto a carrier, to thereby immobilize the antigen; (b) blocking a portion of the carrier surface other than the antigen-adsorbed portion with a protein irrelevant to the antigen; (c) adding a urine sample to be tested and the monoclonal antibody of the present invention so as to bind the monoclonal antibody with the immobilized antigen and free uracil (or thymine) in a competitive manner, thereby forming an immobilized antigen-antibody complex and a free uracil (or thymine)-antibody complex; (d) washing off the free uracil (or thymine)-antibody complex and reacting an enzyme-labeled secondary antibody with the immobilized antigen-antibody complex; and (e) reacting an appropriate substrate with the enzyme and measuring absorbance.

In the aforementioned step (a), the aforementioned immunogens can be utilized as the (5-halogeno-1-carboxymethyluracil)-Schlepper complex serving as an immobilized antigen. Basically, the same immunogen as used to prepare an antibody-producing cell is preferably employed as the immobilized antigen. However, the 5-halogeno-1-carboxymethyluracil-Schlepper complex serving as the immobilized antigen is not necessarily identical with the

same complex serving as the immunogen. Briefly, 5-halogeno-1-carboxymethyluracil-BSA complex can serve as the immunogen and 5-halogeno-1-carboxymethyluracil-KLH can serve as the immobilized antigen.

No particular limitation is imposed on the carrier onto which the antigen is immobilized, and any carrier which is typically used in ELISA can be used. Examples of the carrier include a microplate or beads formed of polystyrene or polyvinyl resin, with a 96-well microplate being preferred. The concentration of the antigen is not particularly limited and is appropriately predetermined generally within a wide range of approximately 0.01-100 $\mu\text{g/mL}$, preferably 0.4-50 $\mu\text{g/mL}$. When a 96-well microplate is used as a carrier, the antigen may be used in a volume sufficient to cover the bottom surfaces of the wells in their entirety. Generally, a volume of approximately 20-100 μL per well is preferably used.

No particular limitation is imposed on the adsorption conditions. Typically, allowing to stand at approximately 4-40°C for approximately one hour to one night is suitable, and allowing to stand at approximately 4°C for approximately one night or at approximately 37°C for approximately two hours is preferred.

In step (b), examples of proteins which can be used for blocking include calf serum, ovalbumin, bovine serum albumin, fetal calf serum, skim milk, and gelatin, with gelatin and calf serum being preferred. No particular limitation is imposed on the blocking conditions, and typically, allowing

to stand at approximately 4-40°C for approximately one hour to one night is suitable, and allowing to stand at approximately 4°C for approximately one night or at approximately 37°C for approximately two hours is preferred. After completion of blocking, the microplate is washed by use of a buffer. The buffer used herein is not particularly limited, and, for example, phosphate buffered saline (PBS) containing Tween 20 (pH 7.3-7.7) is preferred.

In step (c), no particular limitation is imposed on the specific conditions, and typically, allowing to stand at approximately room temperature to 40°C for approximately 0.5-3 hours is suitable, and allowing to stand at approximately 37°C for approximately one hour is preferred. After completion of reaction, the microplate is washed by use of a buffer. The buffer herein used is not particularly limited, and, for example, PBS containing Tween 20 (pH 7.3-7.7) is preferred.

As the secondary antibody used in step (d), an enzyme-labeled anti-mouse immunoglobulin antibody can be used. Examples of the enzyme include alkaline phosphatase (AP), horse radish peroxidase (HRPO), β -galactosidase (β -GS), glucose oxidase (GO), and urease. Among such secondary antibodies, an AP-labeled anti-mouse immunoglobulin antibody is preferred. The secondary antibody is typically diluted for use, and there is preferably used a secondary antibody which has been diluted at a dilution factor of approximately 100-10,000 times, more preferably approximately 200-1,000

times, based on the concentration of the monoclonal antibody (primary antibody) which has been bound with the microplate via the uracil-Schlepper complex. In order to dilute the secondary antibody, a protein used for blocking that is diluted with PBS; e.g., PBS (pH 7.3-7.7) containing 0.1% gelatin, is preferably used. The reaction conditions are not particularly limited, and the reaction is typically carried out at approximately 37°C for approximately one hour. After completion of the reaction, the reaction system is washed with a buffer. Through the above reaction, the secondary antibody is caused to bind with the monoclonal antibody which is bound with the microplate via the immobilized antigen.

In step (e), a substrate is reacted with the enzyme bound to the secondary antibody so as to allow the substrate to develop color. In the reaction system, an optional color developer may further be added. The change in absorbance is measured. More specifically, in a preferred method, when alkaline phosphatase is employed as the labeling enzyme to be bound with the secondary antibody, p-nitrophenylphosphoric acid is employed as a substrate, to thereby develop color through enzymatic reaction. The enzymatic reaction is terminated through addition of 2N NaOH, and absorbance is measured at 415 nm.

In the case in which peroxidase is employed as the labeling enzyme to be bound with the secondary antibody, hydrogen peroxide and o-phenylenediamine are preferably employed as a substrate and a color developer, respectively.

In this case, specific conditions are not particularly limited, and there is used a method in which a solution of a color developer is added to a reaction system; the system is allowed to react at approximately 25°C for approximately 10 minutes; and the enzymatic reaction is terminated through addition of 4N sulfuric acid.

When o-phenylenediamine is employed as a color developer, the absorbance at 492 nm is measured. In addition, sensitizing by use of an avidin-biotin system may also be employed.

In a simple manner for carrying out the aforementioned immunochemical assay, a diagnostic agent for diagnosing DPD deficiency that contains the antiuracil monoclonal antibody of the present invention is used.

Specifically, the diagnostic agent—used for detecting or quantitating uracil or thymine in a sample to be assayed—is a kit containing the monoclonal antibody of the present invention and optional uracil or 5-halogeno-1-carboxymethyluracil-Schlepper complex.

The diagnostic agent may be a set containing, in addition to the monoclonal antibody of the present invention, at least one to five ingredients selected from among carriers for immobilization, labeling agents, substrates (detection agents) corresponding to the labeling agents, antigens, and secondary antibodies (e.g., anti-mouse immunoglobulin). When the set contains a labeling agent, the labeling agent may be conjugated in advance with an arbitrary ingredient such as a

secondary antibody. Examples of the labeling agent include a variety of compounds such as radioisotopes, enzymes, and fluorescent substances. Of these, enzymes are preferred from a variety of viewpoints, such as operability.

The diagnostic agent may further contain, for measurement purposes, appropriate ingredients such as a diluent for an antibody or an antigen, a diluent for reaction, a buffer, a detergent, a substrate-dissolving agent, a reaction-terminating agent, and a solid-adsorption-preventing agent.

Example 1. Preparation of 5-bromo-1-carboxymethyluracil-Schlepper complex

By use of an Immject Immunogen EDC Conjugation kit (product of Pierce) and in accordance with the manual attached thereto, 5-bromo-1-carboxymethyluracil was bound to bovine serum albumin (BSA) or hemocyanin (KLH), to thereby prepare a 5-bromo-1-carboxymethyluracil-BSA complex or a 5-bromo-1-carboxymethyluracil-KLH complex. Specifically, the following method was employed.

- 1) 5-Bromo-1-carboxymethyluracil (4 mg) was dissolved in a conjugation buffer (1 mL; 0.1 M MES (2-(N-Morpholino)-ethane sulfonic acid), 0.9 M NaCl, 0.02%, NaN₃, pH 4.7).

- 2) BSA (or KLH) (10 mg) was dissolved in another portion of the conjugation buffer (1 mL).

- 3) The 5-bromo-1-carboxymethyluracil solution (50 μ L) prepared in step 1) and 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC) (125 μ L; when KLH is used in

place of BSA, 50 μ L) were added to the BSA (or KLH) solution (200 μ L) prepared in step 2).

4) The solution prepared in step 3) was allowed to stand at 37°C for 2 hours, and subjected to centrifugation at 10,000 rpm for 5 minutes, to thereby remove pellets.

5) The solution obtained in step 4) was added to a D-salt dextran desalting column, and the eluate was fractionated at 0.5 mL/tube. The absorbance of each fraction was measured at 280 nm and 260 nm. The fractions corresponding to the first peak were collected, to thereby obtain a solution of 5-bromo-1-carboxymethyluracil-BSA (or KLH) complex.

6) The presence of the complex was confirmed through analysis of absorbance in the vicinity of OD 260 nm, which is attributed to the pyrimidine skeletal structure of 5-bromo-1-carboxymethyluracil. As a result, in relation to the 5-bromo-1-carboxymethyluracil-BSA complex, it was deduced that 7.31 molecules of 5-bromo-1-carboxymethyluracil had been bound to one BSA molecule, whereas in relation to the 5-bromo-1-carboxymethyluracil-KLH complex, it was deduced that 26 molecules of 5-bromo-1-carboxymethyluracil had been bound to one KLH molecule.

The above-described complexes can be utilized to prepare a hybridoma and a monoclonal antibody.

Example 2. Preparation of antibody-producing cells

To each of five BALB/c mice, complete Freund's adjuvant (200 μ L) containing the antigen prepared in Example 1 (5-

bromo-1-carboxyluracil-KLH, 10 μ g) was injected intraperitoneally. Two weeks thereafter, incomplete Freund's adjuvant (200 μ L) containing the same antigen (10 μ g) was intraperitoneally administered to the same mice.

A further 2 weeks thereafter, blood was drawn from the fundus of the mice. While the thus-collected blood was used as an antibody source, inhibition of antigen-antibody reaction by uracil was checked. PBS solution (200 μ L) containing the antigen (10 μ g) was administered intravenously to a mouse whose serum showed the highest level of inhibition by uracil. Three days thereafter, the spleen of the mouse was aseptically removed, and washed with RPMI1640 medium twice. The spleen was transferred to a clean bench, and washed with the same medium two additional times. Subsequently, the spleen was minced in a Petri dish, and the thus-obtained spleen cells were suspended in RPMI1640 medium. The cell suspension was placed in a 15-mL centrifuge tube and subjected to centrifugation for 7 minutes at 1,000 rpm at room temperature. The supernatant was discarded.

The cell sediments were suspended in RPMI1640 medium (10 mL), and the resultant cell suspension was allowed to stand for 3 minutes at room temperature. The cell suspension was withdrawn to a 50-mL centrifuge tube so as not to carry the tissue mass at the bottom of the centrifuge tube, and subjected to centrifugation for 7 minutes at 1,000 rpm at room temperature. The supernatant was discarded, and RPMI1640 medium was added to the centrifuge tube such that

the total volume of the suspension became 10 mL. The number of spleen cells having a nucleus was counted with a hemocytometer. The spleen cells prepared through the above-described procedure were used for fusion with myeloma cells.

Example 3. Cell fusion

(1) Preparation of myeloma cells

P3U1 cells of 8-azaguanine-resistant mouse (BALB/c mouse) myeloma strain were used for cell fusion with the aforementioned spleen cells. The P3U1 cells had been subcultured in advance with RPMI1640 medium supplemented with 10% fetal bovine serum (FBS).

The P3U1 cells of day 1 subculture were transferred into a 50-mL centrifuge tube, and subjected to centrifugation for 5 minutes at 1,000 rpm at room temperature. The supernatant was discarded, and the P3U1 cells were suspended in RPMI1640 medium (10 mL). The number of cells was counted with a hemocytometer. The thus-prepared myeloma cells (P3U1 cells) were used for cell fusion.

(2) Cell fusion

The number of cells of the P3U1 cells prepared in step (1) and that of the sensitized cells prepared in Example 2 were counted, and the cells were mixed at the following ratio (by number of cells). P3U1 cells : sensitized cells = 1 : 5. The mixture was allowed to stand for 10 minutes at room temperature, followed by centrifugation for 5 minutes at 1,000 rpm at room temperature. The supernatant was discarded, and the cell pellet was loosened by tapping the centrifuge

tube. 50% Polyethylene glycol (average molecular weight = 1,000) was added, over 10 seconds, to the centrifuge tube containing the loosened cells in an amount of 1 mL per combined cell count of 2×10^8 cells while spinning the centrifuge tube. Spinning was continued for a further 50 seconds. Subsequently, while spinning, RPMI1640 medium (15 mL) was added over 3 minutes, and an additional 25 mL of the medium was added over 1 minute. The contents of the centrifuge tube were made uniform through gentle pipetting, followed by centrifugation for 5 minutes at 1,000 rpm at room temperature. The supernatant was discarded, and the pellet was suspended in HAT medium supplemented with 10% fetal bovine serum (to make the concentration of the sensitized cells 1.6×10^8 cells/mL). The thus-prepared cell suspension was plated in a 96-well flat-bottomed plate at 200-250 μ L/well. The cells were incubated for a period of 10 to 14 days at 37°C under 5% CO₂.

Example 4. Screening of hybridoma

The hybridoma cells obtained in Example 3 were subjected to a screening test through ELISA.

Into a 96-well immunoplate, uracil-BSA complex (10 μ g/mL as reduced to BSA) was placed at 50 μ L/well, and allowed to stand overnight at 4°C. Subsequently, the immunoplate was washed 7 times with PBS containing 0.1% polyoxyethylene(20) sorbitan monolaurate (product of Wako Pure Chemical Industries, Ltd.; a product equivalent to Tween 20) (pH 7.3-7.7, product of Nissui Chemical; hereinafter

referred to as PBS-T). Subsequently, 0.2% gelatin (200 μ L) was added to each well, and the immunoplate was left for 2 hours at 37°C, or overnight at 4°C.

Subsequently, the immunoplate was washed five times with PBS-T, and one of the below-described samples (50 μ L/well) was added to the immunoplate, which was then left for 1 hour at 37°C.

(i) a reaction mixture prepared by mixing the supernatant of the hybridoma culture obtained in Example 3 (hereinafter referred to as the hybridoma culture supernatant) (30 μ L) and PBS (30 μ L) for reaction at 37°C for one hour;

(ii) a reaction mixture obtained by mixing the hybridoma culture supernatant (30 μ L) and a solution (30 μ L) of uracil in PBS (uracil concentration: 1 mg/mL) for reaction at 37°C for one hour; and

(iii) 1 μ g/mL healthy mouse IgG (product of Biological). Subsequently, the wells were washed five times with PBS-T, and to each well was added alkaline-phosphatase-labeled (AP-labeled) anti-mouse-multivalent-immunoglobulin antibody (product of Sigma), which had been 1,000-fold diluted with 0.1% gelatin, (50 μ L). Subsequently, the mixture was allowed to stand for one hour at 37°C. Subsequently, the wells were washed five times with PBS-T. A solution of disodium p-nitrophenylphosphate (product of Wako Chemical Industries, Ltd.) dissolved in substrate buffer (1 mg/mL) was added to each well (100 μ L/well), and left for 30 minutes at 37°C. To

the reaction mixtures, 2N NaOH (50 μ L/well) was added and then mixed by use of a platemixer, to thereby stop the reaction. Absorbance at 415 nm was measured by use of a microplate reader.

The percent absorption was calculated by use of the following equation:

$$\text{Percent Absorption (\%)} = \left(1 - \frac{\text{OD}_{\text{uracil}} - \text{OD}_{\text{control}}}{\text{OD}_{\text{PBS}} - \text{OD}_{\text{control}}} \right) \times 100$$

wherein,

OD_{PBS}: Absorbance measured by means of ELISA for a sample in which the hybridoma culture supernatant had been pre-treated only with PBS),

OD_{uracil}: Absorbance measured by means of ELISA for a sample in which the hybridoma culture supernatant had been pre-treated with uracil in PBS), and

OD_{control}: Absorbance measured in ELISA of a sample treated only with healthy mouse IgG).

Example 5. Cloning

The monoclonal-antibody-producing hybridoma obtained in Example 4 was cloned twice through a known customary limiting dilution method.

Specifically, the number of the monoclonal-antibody-producing hybridoma cells obtained in Example 4 was counted. A hybridoma cell and RPMI1640 (0.2 mL) containing 10% FBS and 10% BM condimed H1 (Registered Trademark of Boehringer Mannheim) were placed into each well of an immunoplate, and incubated under 5% CO₂ at 37°C for 10-14 days.

The resultant hybridoma clones were subjected to screening through ELISA as described in Example 4. Subsequently, the selected hybridoma clones were cloned again through the above-described method, followed by screening through the aforementioned ELISA again. As a result, the cloned cells (2D3 and 2H10) which produce an antibody specific to uracil and thymine were screened out from the monoclonal-antibody-producing hybridomas. One of the thus-obtained cloned cells (2D3), mouse hybridoma CMU-1, was deposited with the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, Ministry of International Trade and Industry under the Budapest Treaty, (accession No.: FERM BP-6870).

Example 6. Storage of anti-uracil-monoclonal-antibody-producing hybridomas

The monoclonal-antibody-producing hybridoma clones obtained in Example 5 were added to RPMI1640 medium containing 10% FBS and 10% DMSO so that the hybridoma concentration became 1×10^7 cells/mL. Aliquots (1 mL) of the resultant mixture were placed in CryoTubes (product of Nunc), followed by freezing in a deep freezer at -90°C , and storing in liquid nitrogen.

Example 7. Preparation of monoclonal antibody and its evaluation

The monoclonal-antibody-producing hybridoma, mouse hybridoma CMU-1, obtained in Example 5 was cultured in RPMI1640 medium containing 10% FBS and 10% BM conditioned H1

(product of Boehringer Mannheim) under 5% CO₂ at 37°C for 2-3 days.

By use of the hybridoma culture supernatant obtained from the above-described incubation, the reactivity of the monoclonal antibody, CMU-1, of the present invention was studied through the indirect competitive inhibition ELISA described in Example 4.

Specifically, an aliquot (50 µL) of uracil-BSA complex (10 µg/mL) was placed in each well of a 96-well immunoplate (Immunoplate I, product of Nunc), and allowed to stand overnight at 4°C. Each well was washed five times with PBS-T (200 µL) containing 0.1% Tween 20. Subsequently, to each well of the immunoplate, PBS (200 µL) containing 0.2% gelatin serving as a blocking agent was added, and the mixture was allowed to stand overnight at 4°C. Each well was again washed five times with PBS-T, to thereby prepare a plate.

Meanwhile, each of the follow samples:

(i) a mixture of the aforementioned hybridoma culture supernatant (30 µL) and PBS (30 µL);

(ii) a reaction mixture obtained by mixing the hybridoma culture supernatant (30 µL) and PBS (30 µL) containing uracil at a concentration of 0.03-1.0 mg/mL and causing reaction to proceed at 37°C for one hour; and

(iii) 1 µL/mL healthy mouse IgG (product of Biologicals) was added to a 96-well plate to which the above antigen had been adsorbed (50 µL/well), and the plate was left to stand for one hour at 37°C. Subsequently, the plate

was washed five times with PBS-T, and to each well, aliquots (50 μ L) of AP-labeled anti-mouse antibody (serving as the second antibody, product of Sigma), which had been 1,000-fold diluted with PBS containing 0.1% gelatin, were added, and the mixtures were allowed to stand for one hour at 37°C. Subsequently, each well was washed five times with PBS-T. An enzyme substrate buffer solution (p-nitrophenylphosphate, 1 mg/mL, pH 9.8) was added to each well (100 μ L/well), and left to stand for 30 minutes at 37°C. To the reaction mixtures, 2N NaOH (50 μ L/well) was added, to thereby stop the reaction. Absorbance at 415 nm was measured by use of a microplate reader. For comparison, the same procedure was independently performed on hybridoma FERM BP-6141 described in PCT publication No. WO99/20748. The results are shown in Fig. 1. The percent absorption was calculated from the above-described equation.

As shown in Fig. 1, the monoclonal antibody of the present invention enables detection of uracil in a concentration range of 0.005-0.5 mg/mL. The detection sensitivity for uracil detection is considerably enhanced as compared with monoclonal antibody SU-1 originating from hybridoma FERM BP-6141.

Example 8. Cross reactivity test

In a manner similar to that of Example 7, reactivity of the monoclonal antibody of the present invention with other compounds was investigated. Specifically, the procedure of step (ii) of Example 7 for investigating inhibition with

uracil was repeated, except that pseudouridine, dihydrouracil, dihydrothymine, thymine, cytosine, or N-carbamyl- β -alanine (1 mg/mL) was used instead of uracil (1 mg/mL), to thereby investigate reactivity of the monoclonal antibody of the present invention with uracil compounds. As a comparative antibody, a monoclonal antibody (SU-1) derived from hybridoma "FERM BP-6141" described in WO 99/20748 was used. Table 1 shows the results.

Table 1

Compound*	Monoclonal antibody	
	Present invention (CMU-1)	Comparative (SU-1)
Uracil	95-96%	41%
Thymine	91-97%	5%
N-Carbamyl- β -alanine	8%	91%
Dihydrouracil	8%	2%
Dihydrothymine	22%	—
Pseudouridine	32%	26%
Cytosine	15%	15%

*Final concentration 0.5 mg/mL

As shown in Table 1, the monoclonal antibody of the present invention reacted strongly with uracil and thymine and exhibited dose-dependent response. However, reactivity thereof with N-carbamyl- β -alanine, dihydrouracil, dihydrothymine, pseudouridine, and cytosine was found to be low. In contrast, the comparative antibody exhibited higher reactivity with N-carbamyl- β -alanine than with uracil or thymine.

In addition, the antibody of the present invention exhibited no dose-dependent response to N-carbamyl- β -alanine,

dihydrouracil, dihydrothymine, and cytosine, but did exhibit dose-dependent response to pseudouridine. Thus, the concentration-percent absorption relationship in terms of uracil, thymine, and pseudouridine to which dose-dependent response had been confirmed was investigated. Fig. 2 shows the results, and Table 2 shows the concentrations (mg/mL) of respective compounds when the percent absorption reached 30% (ED₃₀).

Table 2

Compound	ED ₃₀ (mg/mL)
Uracil	0.03
Thymine	0.015
Pseudouridine	0.39

The results indicated that the concentration of uracil was 1/13 that of pseudouridine so as to exhibit the same percent absorption and that the concentration of thymine was 1/26 that of pseudouridine so as to exhibit the same percent absorption. Accordingly, the binding ability of uracil was found to be 13 times that of pseudouridine and the binding ability of thymine was found to be 26 times that of pseudouridine.

Industrial Applicability

The monoclonal antibody of the present invention is able to selectively assay, at high speed and in a simple manner, uracil and thymine contained in a sample. In addition, the monoclonal antibody is useful for diagnosing

DPD deficiency by use of a human urine sample. Particularly, the monoclonal antibody is considerably useful for screening DPD-deficient cancer patients with contraindication to the administration of a pyrimidine fluoride-based antitumor agent.

Claims

1. A monoclonal antibody, characterized by reacting strongly with uracil and thymine but scarcely with N-carbamyl- β -alanine.
2. A monoclonal antibody as described in claim 1, which exhibits scarce or low reactivity with pseudouridine, dihydrouracil, and dihydrothymine.
3. A monoclonal antibody as described in claim 1, which exhibits a selectivity in cross reaction with N-carbamyl- β -alanine of 10% or less, when the selectivity in cross reaction with uracil or thymine is 90% or more.
4. A monoclonal antibody as described in claim 1 or 3, which exhibits a selectivity in cross reaction with N-carbamyl- β -alanine of 10% or less; a selectivity in cross reaction with pseudouridine of 33% or less; a selectivity in cross reaction with dihydrouracil of 8% or less; and a selectivity in cross reaction with dihydrothymine of 23% or less; when the selectivity in cross reaction with uracil or thymine is 90% or more.
5. A monoclonal antibody as described in any one of claims 1 to 4, which is produced from a hybridoma which is formed from a myeloma cell and an antibody-producing cell derived from an animal to which 5-halogeno-1-carboxymethyluracil has been administered.
6. A monoclonal antibody as described in claim 5, wherein the hybridoma is FERM BP-6870 strain.
7. A hybridoma producing a monoclonal antibody as

recited in any one of claims 1 to 6.

8. A method for immunochemically assaying uracil and thymine by use of a monoclonal antibody as recited in any one of claims 1 to 6.

9. A diagnostic agent for diagnosing DPD deficiency, which agent contains a monoclonal antibody as recited in any one of claims 1 to 6.

10. A method for diagnosing DPD deficiency, characterized by assaying uracil and thymine in a specimen by use of a diagnostic agent for diagnosing DPD deficiency as recited in claim 9.

11. Use of a monoclonal antibody as recited in any one of claims 1 to 6 in order to produce a diagnostic agent for diagnosing DPD deficiency.

Abstract

A monoclonal antibody which reacts strongly with uracil and thymine but scarcely with N-carbamyl- β -alanine; a hybridoma producing this monoclonal antibody; a method of immunochemically assaying uracil or thymine characterized by using the above-described monoclonal antibody; and diagnostics for DPD deficiency containing the above monoclonal antibody. Because of high sensitivity and specific reaction with uracil and thymine, the above-described monoclonal antibody enables convenient, quick, and selective assaying of uracil and thymine in a sample. The antibody is useful in screening patients with DPD deficient cancer with contraindication to the administration of pyrimidine fluoride-based antitumor agents.

Fig. 1

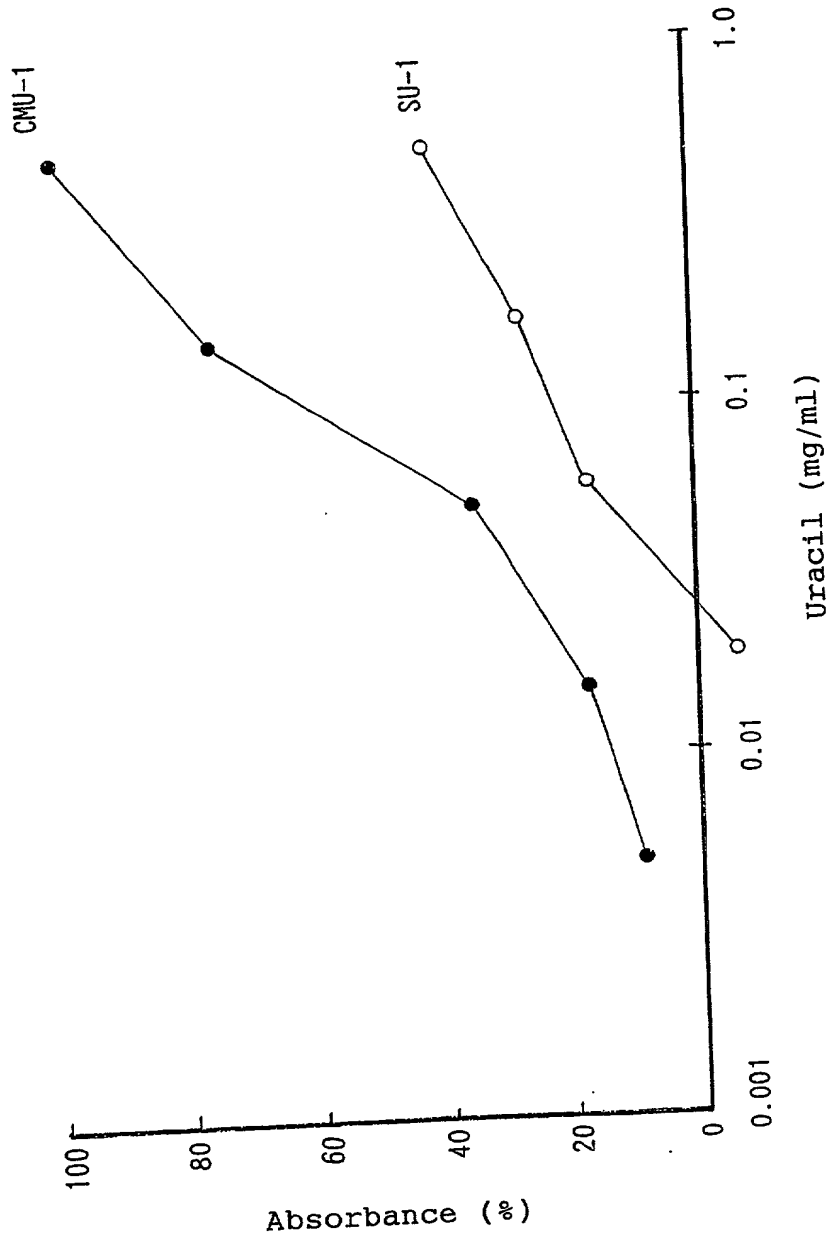
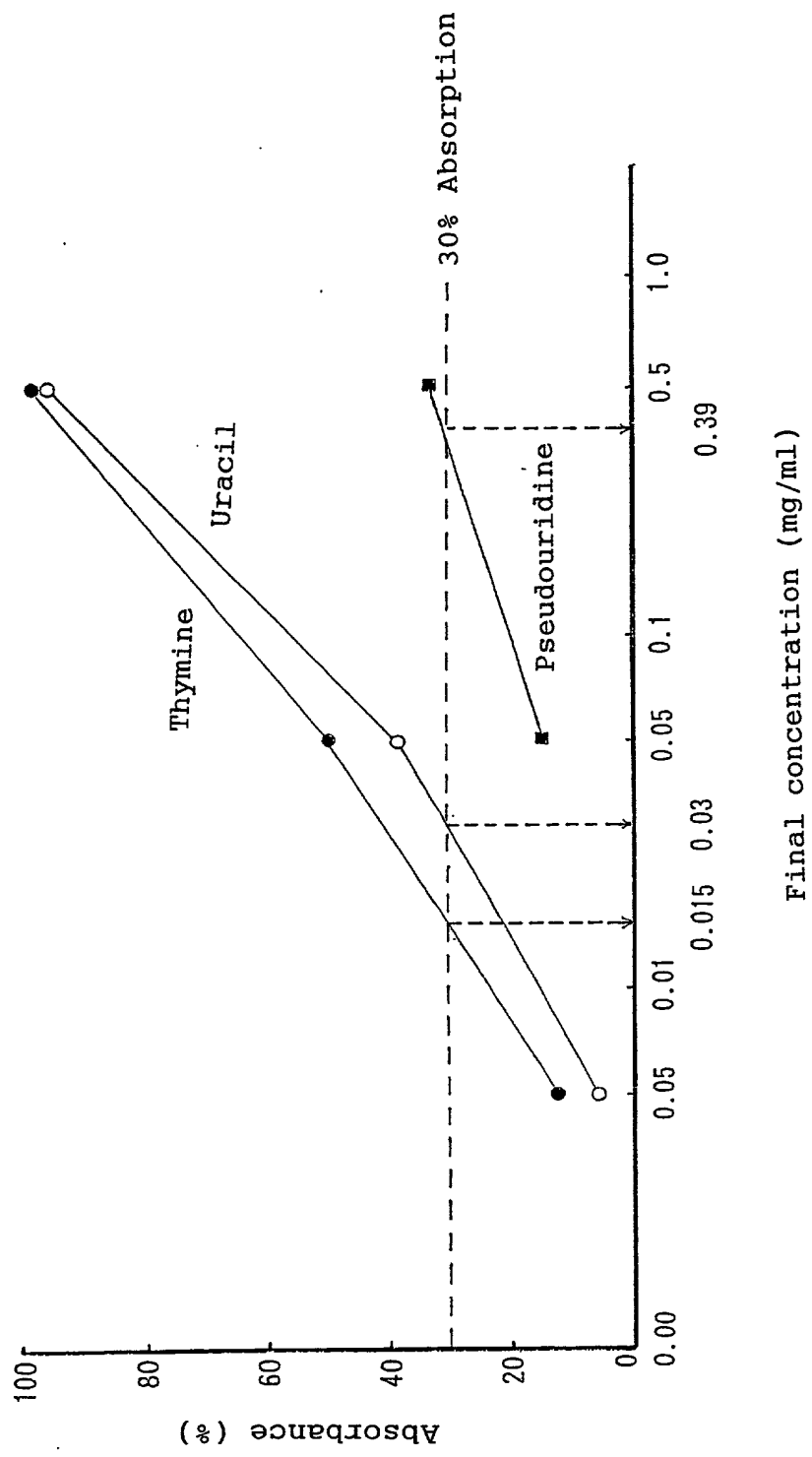


Fig. 2



Declaration and Power of Attorney For Patent Application

特許出願宣言書及び委任状

Japanese Language Declaration

日本語宣言書

下記の氏名の発明者として、私は以下の通り宣言します。

As a below named inventor, I hereby declare that:

私の住所、私書箱、国籍は下記の私の氏名の後に記載された通りです。

My residence, post office address and citizenship are as stated next to my name.

下記の名称の発明に関して請求範囲に記載され、特許出願している発明内容について、私が最初かつ唯一の発明者（下記の氏名が一つの場合）もしくは最初かつ共同発明者（下記の名称が複数の場合）であると信じています。

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled.

抗ウラシルモノクローナル抗体

ANTIURACIL MONOCLONAL ANTIBODY

上記発明の明細書は、

the specification of which

☐ 本書に添付されています。

☐ is attached hereto.

☒ 2000年10月18日に提出され、米国出願番号は特許協定条約国際出願番号を PCT/JP00/07216 とし、
(該当する場合) _____ に訂正されました。

☒ was filed on October 18, 2000
as United States Application Number or
PCT International Application Number
PCT/JP00/07216 and was amended on
_____ (if applicable).

私は、特許請求範囲を含む上記訂正後の明細書を検討し、内容を理解していることをここに表明します。

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

私は、連邦規則法典第37編第1条56項に定義されるとおり、特許資格の有無について重要な情報を開示する義務があることを認めます。

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56.

Japanese Language Declaration

(日本語宣言書)

私は、米国法典第35編119条 (a) - (d) 項又は365条 (b) 項に基づき下記の、米国以外の国の少なくとも一カ国を指定している特許協力条約365 (a) 項に基づく国際出願、又は外国での特許出願もしくは発明者証の出願についての外国優先権をここに主張するとともに、優先権を主張している、本出願の前に出願された特許または発明者証の外国出願を以下に、枠内をマークすることで、示しています。

Prior Foreign Application(s)

外国での先行出願
11-296817

Japan

(Number)
(番号)

(Country)
(国名)

(Number)
(番号)

(Country)
(国名)

私は、第35編米国法典119条 (e) 項に基づいて下記の米国特許出願規定に記載された権利をここに主張いたします。

(Application No.)
(出願番号)

(Filing Date)
(出願日)

私は、下記の米国法典第35編120条に基づいて下記の米国特許出願に記載された権利、又は米国を指定している特許協力条約365条 (c) に基づく権利をここに主張します。また、本出願の各請求範囲の内容が米国法典第35編112条第1項又は特許協力条約で規定された方法で先行する米国特許出願に開示されていない限り、その先行米国出願書提出日以降で本出願書の日本国内または特許協力条約国際提出日までの期間中に入手された、連邦規則法典第37編1条56項で定義された特許資格の有無に関する重要な情報について開示義務があることを認識しています。

(Application No.)
(出願番号)

(Filing Date)
(出願日)

(Application No.)
(出願番号)

(Filing Date)
(出願日)

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I hereby claim foreign priority under Title 35, United States Code, Section 119 (a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate, or Section 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed.

Priority Claimed

優先権主張

19/10/1999

(Day/Month/Year Filed)
(出願年月日)

☒

Yes

はい

☐

No

いいえ

(Day/Month/Year Filed)
(出願年月日)

☐

Yes

はい

☐

No

いいえ

I hereby claim the benefit under Title 35, United States Code, Section 119(e) of any United States provisional application(s) listed below.

(Application No.)
(出願番号)

(Filing Date)
(出願日)

I hereby claim the benefit under Title 35, United States Code, Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code Section 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of application.

(Status: Patented, Pending, Abandoned)
(現況: 特許許可済、係属中、放棄済)

(Status: Patented, Pending, Abandoned)
(現況: 特許許可済、係属中、放棄済)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

(日本語宣言書)

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith: *(list name and registration number)*



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(第三以降の共同発明者についても同様に記載し、署名すること)

(Supply similar information and signature for third and subsequent joint inventors.)